ALLOZYME VARIATION AND POPULATION SUBSTRUCTURING IN THE CAUCASIAN GROUND LIZARDS

Lacerta derjugini AND Lacerta praticola

Ross D. MacCulloch,^{1,2} Ilya S. Darevsky,³ Robert W. Murphy,¹ and Jinzhong Fu¹

Submitted August 11, 1997.

Genetic diversity at 35 allozyme loci was surveyed in *Lacerta derjugini* (3 populations) and *L. praticola* (2 populations). Indices of variability were consistent with those found in other Caucasian *Lacerta*. There was little genetic substructuring between two populations of *L. praticola* despite considerable geographic separation. Conversely, populations of *L. derjugini* in close proximity to one another exhibited considerable substructuring.

Key words: Lacerta, population genetics, protein electrophoresis, allozymes.

INTRODUCTION

The "ground lizards" Lacerta derjugini Bischoff and L. praticola Lantz et Cyrén, are interesting components of the Caucasus lizard fauna. The preferred habitat of these two species is ground vegetation, in either forested or open areas, whereas other small Caucasian Lacerta occupy rocky habitats. For this reason, among others, the two species have not been considered members of the speciose "rock lizard" group (Darevsky, 1967). Later investigations, however, found a close relationship among these two species and other Caucasian lacertas of the saxicola group (Arnold, 1989; Fu et al., in press; Grechko et al., 1993; Mayer and Lutz, 1989; Murphy et al., 1996a).

Recently, a preliminary phylogeny of Caucasian *Lacerta* based on allozymes found inconsistencies at several nodes (Murphy et al., 1996a). Some of these inconsistencies were due to the uncertainty of the phylogenetic relationships of the two ground lizard

MATERIAL AND METHODS

Specimens of *Lacerta derjugini orlowae* were collected from Achaldaba, Bakuriani, and Zedazeni, Georgia. *Lacerta praticola pontica* were collected from Sochi and Tul'skaya, Krasnodar District, Russia. Specimens were euthanised by an overdose of sodium pentobarbital and dissected immediately following euthanasia. Liver, heart and skeletal muscle were removed and frozen in liquid nitrogen. Specimens are deposited in the herpetological collection of the Royal Ontario Museum (ROM); collection details and specimen numbers are available upon request.

Enzymes were separated by horizontal starch gel electrophoresis. All procedures, protocols and allelic nomenclature follow Murphy et al. (1996b). The analysis utilized 27 enzyme systems encoded by 35 presumptive loci. Wherever possible, gene products were resolved on two buffer systems to maximize expression of all variants. Enzyme names, EC numbers and specific buffer systems for the separation of locus

species *L. derjugini* and *L. praticola*. These two species must therefore be examined more closely in order to solidify their place in the evolutionary history of *Lacerta*. The first step toward this goal is a population-level allozyme assay of a broad geographic representation of a number of individuals of these species.

Centre for Biodiversity and Conservation Biology, Royal Ontario Museum, 100 Queen's Park, Toronto, Ontario M5S 2C6, Canada.

Address correspondence and reprint requests to Ross MacCulloch, Centre for Biodiversity and Conservation Biology, Royal Ontario Museum, 100 Queen's Park, Toronto, Ontario M5S 2C6, Canada. Phone: +1 (416) 586-5759, Fax: +1 (416) 586-5553. E-mail: rossm@rom.on.ca.

³ Zoological Institute, Russian Academy of Sciences, 1, Universitetskaya nab., St. Petersburg 119034, Russia.

products were the same as those used by Bobyn et al. (1996), Fu et al. (1995) and MacCulloch et al. (1995).

Allozyme data were analyzed using BIOSYS-1 release 1.7 (Swofford and Selander, 1989). The populations were treated separately for analyses of population variability. All loci were evaluated for three parameters of genetic polymorphism: heterozygosity (MHD), number of alleles per locus (MNA) and percentage of loci which were polymorphic (PLP), as well as conformity to Hardy-Weinberg expectations using Levene's (1949) correction for small sample sizes. The populations of each species were examined together for genetic structuring using Wright's (1978) *F*-statistics. Although only one specimen of *L. derjugini* was collected at Zedazeni, it was included in the analysis.

 TABLE 1. Genotype Frequencies for the Polymorphic Loci in

 Lacerta derjugini

Locus	Population		
	Achaldaba	Bakuriani	Zedazeni
sAat-A	aa (10)	aa (8)	ab (1)
	ab (1)	ab (2)	
sAcoh-A	aa (10)	aa (6)	aa (1)
	ab (1)	ab (4)	
Cat-A	aa (7)	aa (7)	aa (1)
	ab (3)	ab (3)	
	ac (1)		
Ck-C	aa (8)	aa (10)	aa (1)
	ab (3)		
Gpi-A	aa (11)	aa (7)	aa (1)
		ab (3)	
sIdh-A	aa (11)	aa (9)	aa (1)
		ab (1)	
Pep-B	aa (11)	aa (8)	aa (1)
		ab (2)	
np-A	aa (7)	aa (9)	aa (1)
	ab (3)	ab (1)	
	bb (1)		
N ¹	11	10	1
MHD ²	0.033 ± 0.016	0.044 ± 0.017	0.028 ± 0.028
MNA ³	1.17 ± 0.07	1.19 ± 0.07	1.03 ± 0.03
PLP^4	8.33	19,44	2.78

Number of individuals examined.

RESULTS

Lacerta derjugini

All individuals from all three populations expressed homozygous allelic products at 27 of the 35 loci: mAat-A, mAcoh-A, Acp-B, Ada-A, Cbp-1, Ck-A, Est-D, \(\beta\)Ga-1, Gcdh-A, Gda-A, \(\beta\)Glur-A, \(\beta\)Glus-A, Gpi-B, G6pdh-A, mIdh-A, Ldh-A, Ldh-B, mMdh-A, sMdh-A, mMdhp-A, sMdhp-A, Mpi-A, Pep-A, Pgm-A, mSod-A, sSod-A and Tpi-A. Genotype frequencies for the eight polymorphic loci are shown in Table 1. Only one locus (sAat-A) was polymorphic in all three populations. At four of the eight polymorphic loci, only one population exhibited polymorphism. In many cases polymorphism resulted from the single appearance of a rare allele. At four of the five loci which were polymorphic in more than one population, all alleles were present in all polymorphic populations. The sole exception was Cat-A, where Cat-A (c) made a single appearance in the Achaldaba population. The indices of genetic variability are shown at the bottom of Table 1. The analysis of L. derjugini included a single individual from Zedazeni. While it is unusual to use a sample size of one for a study of genetic variability, this individual was included because discrete populations often exhibit rare alleles, although in this instance none was found.

Allele frequencies at one locus failed to conform to Hardy-Weinberg probabilities. The Achaldaba population exhibited a slight heterozygote excess at Pnp-A.

The F-statistics for the three populations are as follows: $F_{\rm IS} = -0.254$, $F_{\rm IT} = 0.203$, $F_{\rm ST} = 0.365$. The negative value of $F_{\rm IS}$ is indicative of an intrapopulational heterozygote excess, while the positive value of $F_{\rm IT}$ indicates an interpopulational heterozygote deficiency smaller than the intrapopulational excess. The relatively high positive value of $F_{\rm ST}$ suggests that the three populations do not form a pannictic group.

Lacerta praticola

All individuals in both populations of L. praticola were monoallelic for 20 of the 35 loci: mAat-A, mAcoh-A, Acp-B, Ada-A, Cbp-1, Ck-A, Ck-C, Est-D, β Ga-1, Gcdh-A, Gda-A, β Glur-A, β Glus-A, Gpi-B, mMdh-A, mMdhp-A, Pgm-A, mSod-A, sSod-A and Tpi-A. Genotype frequencies for the 15 polymorphic loci are shown in Table 2. In most cases polymorphism was due to rare alleles. Of the 15 vari-

² Mean heterozygosity by direct count.

³ Mean number of alleles per locus.

⁴ Percentage of loci polymorphic (0.95 criterion).

able loci, only four (sAat-A, Cat-A, Gpi-A, Pep-A) were polymorphic in both populations. At all four of these loci, the Sochi population exhibited a rare allele which was not detected in the sample from Tul'skaya. The Sochi population also exhibited polymorphism at nine loci which were monomorphic in the Tul'skaya population, whereas the Tul'skaya population exhibited polymorphism at only two loci which were invariant at Sochi. The difference in variability between the two populations may have resulted from the unequal sample sizes.

All allele frequencies conformed to Hardy-Weinberg equilibrium with the exception of Pep-B in the Sochi population, which exhibited a heterozygote deficiency. Indices of genetic variability are shown at the bottom of Table 2.

The F-statistics for the two populations are: $F_{\rm IS} = -0.002$, $F_{\rm IT} = 0.023$, $F_{\rm ST} = 0.025$. These values are the lowest of any so far reported in Caucasian Lac-erta. $F_{\rm IS}$ measures intrapopulational heterozygote deficiency or excess; it is close to zero. The positive value of $F_{\rm IT}$ indicates a small interpopulational heterozygote deficiency. The very low positive value of $F_{\rm ST}$ suggests that the two populations could form a panmictic group.

DISCUSSION

Values of MHD, MNA, and PLP of *L. derjugini* and *L. praticola* approximate those of other Caucasian *Lacerta* (*L. caucasica* and *L. daghestanica*, Fu et al., 1995; *L. raddei* and *L. nairensis*, Bobyn et al., 1996; *L. portschinskii*, *L. rudis*, and *L. valentini*, MacCulloch et al., 1995, in press). Heterozygosity in *L. derjugini* and *L. praticola* are similar to the values found in *Podarcis sicula* and *P. melisellensis* from large island and mainland populations (Gorman et al., 1975). Much higher levels of heterozygosity were found in *L. lepida* (Busack, 1987) and in the teiid lizard species *Cnemidophorus tigris* (Gorman et al., 1977).

A study of 10 genera found mean heterozygosity of 0.01 in fossorial lizards, 0.05 in "sit and wait" species and 0.09 in vagile species (Gorman et al., 1977). Heterozygosity in the two species examined in this study resembles that of "sit and wait" lizards, as did heterozygosity values in other bisexual Caucasian *Lacerta* (Bobyn et al., 1996; Fu et al., 1995; MacCulloch et al., 1995; in press). A more extensive sum-

mary reported a mean heterozygosity of 0.051 from 71 lizard taxa or populations, most of whom are vagile or "sit-and-wait" (Sattler and Ries, 1995). This number is also similar to that found in Caucasian *Lacerta*.

TABLE 2. Genotype Frequencies for the Polymorphic Loci in Lacerta praticola

Locus	Population		
Locus	Sochi	Tul'skaya	
sAat-A	aa (38)	aa (19)	
	ab (3)	ab (2)	
	ac (2)		
sAcoh-A	aa (42)	aa (21)	
	ab (1)		
Cat-A	aa (32)	aa (19)	
	ab (4)	ac (2)	
	bb (1)		
	ac (6)		
Gpi-A	aa (38)	aa (19)	
	ab (3)	ac (2)	
	ac (2)		
G6pdh-A	aa (42)	aa (21)	
	ab (1)		
s-Idh-A	aa (41)	aa (21)	
	ab (2)		
m-Idh-A	aa (43)	aa (19)	
		ab (2)	
Ldh-A	aa (42)	aa (21)	
	ab (1)		
Ldh-B	aa (41)	aa (21)	
	ab (2)		
s-Mdh-A	aa (40)	aa (21)	
	ab (3)		
s-Mdhp-A	aa (42)	aa (21)	
	ab (1)		
Mpi-A	aa (41)	aa (21)	
	ab (2)	4.0	
Pep-A	aa (41)	aa (16)	
	ab (2)	ac (5)	
Pep-B	aa (39)	aa (21)	
	ab (3)		
	bb (1)	(10)	
Pnp-A	aa (43)	aa (19)	
		ab (2)	
N	43	21	
MHD	0.025 ± 0.009	0.015 ± 0.008	
MNA	1.46 ± 0.11	1.11 ± 0.05	
PLP	11.43	2.86	

Note. For designations see Table 1.

118 Ross D. MacCulloch et al.

In our study the percentage of loci exhibiting polymorphism was calculated using the 0.95 criterion in BIOSYS-1. Since some studies did not use this criterion, PLP was recalculated without the criterion to facilitate comparison. This produced a different value in the Achaldaba sample of *L. derjugini* (13.89%) and in both samples of *L. praticola* (37.14 and 11.43% at Sochi and Tul'skaya, respectively); the other populations did not change. Other Caucasian *Lacerta* exhibited similar values. Mainland populations of *P. sicula* had PLP of 27 – 45% (Gorman et al., 1975), while *L. lepida* had PLP of 23.1 and 38.5% (Busack, 1987). PLP varied from 10 – 21% in three species of bisexual *Cnemidophorus* (Dessauer and Cole, 1984).

The range of L. praticola is much more extensive than that of L. derjugini. Range size has been shown to be positively correlated with the amount of genetic variability in lacertids (Gorman et al., 1975; MacCulloch et al., in press). The same correlation is apparent in this study.

Comparison of the values of F_{ST} of the two species shows that L. derjugini has a greater value than does L. praticola (0.365 vs. 0.025). F_{ST} is an index of genetic substructuring among populations; the higher the value of F_{ST} the less likelihood that the populations constitute a panmictic group. Comparison of the relative values of F_{ST} in the two species to the distances between sampling sites provides insight into population substructuring. Even though the two sampling locations of L. praticola are separated by more than 100 km and a mountain range, F_{ST} in L. praticola differs little from zero. This very low value, lower than in any other Caucasian Lacerta (Bobyn et al., 1996; Fu et al., 1995; MacCulloch et al., 1995, in press), suggests that gene flow among populations of this species remains high despite geographic obstacles. The samples of L. derjugini were approximately the same distance apart, but there were no obvious physical barriers separating them. Rather, because the three locations were all from within the same river valley system, gene flow should be facilitated among local populations. Despite this, L. derjugini exhibited a much higher F_{ST} than did L. praticola. Because two of the sampling locations for L. derjugini, Achaldaba and Bakuriani, are only some 20 km apart, F_{ST} was recalculated for these two samples alone. The resulting value of 0.288, although lower than that calculated from three populations, is still one order of magnitude greater than F_{ST} in L. praticola and is greater than $F_{\rm ST}$ in any other Caucasian *Lacerta* from within a contiguous distribution. This suggests either the presence of an unreported barrier(s) to gene flow in *L. derjugini* along the Kura river valley system, or some other mechanism which produces genetic substructuring in the species.

While *L. derjugini* and *L. praticola* share the "ground lizard" microhabitat preference, this niche may have been independently derived in each species from the typical rocky habitat preferred by most small species of Caucasian *Lacerta*. This likelihood is supported by the two species' evolutionary relationships. Although the two species are grouped phylogenetically with the Caucasian rock lizards, they were not resolved as sister taxa (Fu et al., in press; Murphy et al., 1996a).

ACKNOWLEDGMENTS

We are grateful to B. Tuniev and M. Bakradze for specimen collection. Laboratory assistance was provided by A. Misir. All collecting and euthanasia were performed under approved animal protocols. Import permits were issued by Agriculture Canada, This study was supported by Natural Sciences and Engineering Council of Canada Grant No. A3148 and the ROM Department of Museum Volunteers (RWM), by the ROM Foundation (RDM, RWM), and by the Zoological Institute of the Russian Academy of Sciences, the Russian Scientific and Technical Program "Priority Trends in Genetics," the International Science Foundation (No. J3Y100) and the Russian Foundation for Basic Research (ISD). This is contribution number 95 from the Centre for Biodiversity and Conservation Biology of the Royal Ontario Museum.

REFERENCES

Arnold E. N. (1989), "Towards a phylogeny and biogeography of the Lacertidae: relationships within an old-world family of lizards derived from morphology," *Bull. Br. Mus. Nat. Hist.* (Zool.), **55**, 209 – 257.

Bobyn M. L., Darevsky I. S., Kupriyanova L. A., Mac-Culloch R. D., Upton D. E., Danielyan F. D., and Murphy R. W. (1996), "Allozyme variation in populations of *Lacerta raddei* and *Lacerta nairensis* (Sauria: Lacertidae) from Armenia," *Amphibia-Reptilia*, 17, 233 – 246.

Busack S. D. (1987), "Morphological and biochemical differentiation in Spanish and Moroccan populations of the lizard, *Lacerta lepida*," *J. Herpetol.*, **21**, 277 – 284.

- Darevsky I. S. (1967), Rock Lizards of the Caucasus: Systematics, Ecology, and Phylogenesis of the Polymorphic Groups of Caucasian Rock Lizards of the Subgenus Archaeolacerta, Nauka, Leningrad [in Russian].
- **Dessauer H. C. and Cole C. J.** (1984), "Influence of gene dosage on electrophoretic phenotypes of proteins from lizards of the genus *Cnemidophorus*," *Comp. Biochem. Physiol.*, **77B**, 181 189.
- Fu J., Darevsky I. S., MacCulloch R. D., Kupriyanova L. A., Roytberg E. S., Sokolova T. M., and Murphy R. W. (1995), "Genetic and morphological differentiation among Caucasian rock lizards of the *Lacerta caucasica* complex," *Russ. J. Herpetol.*, 2, 36 43.
- **Fu J., Murphy R. W., and Darevsky I. S.** (1997), "Toward the phylogeny of Caucasian rock lizards: Implications from mitochondrial DNA gene sequences (Reptilia: Lacertidae)," *Zool. J. Linn. Soc*, **122**, 463 477.
- Gorman G. C., Soulé M., Yang S. Y., and Nevo E. (1975), "Evolutionary genetics of insular Adriatic lizards," *Evolution*, **29**, 52 71.
- Gorman G. C., Kim Y. J., and Taylor C. E. (1977), "Genetic variation in irradiated and control populations of *Cnemidophorus tigris* (Sauria, Teiidae) from Mercury, Nevada, with a discussion of genetic variability in lizards," *Theor. Appl. Genet.*, 49, 9 14.
- Grechko V. V., Kataev M. V., Mel'nikova M. N., and Darevsky I. S. (1993), "Relatedness of parthenogenetic *Lacerta* lizards and their presumable parental bisexual species deduced from comparative data on amplification products obtained by single-primer polymerase chain reaction," *Mol. Biol.*, 27, 891 897.
- **Levene H.** (1949), "On a matching problem arising in genetics," *Ann. Math. Stat.*, **20**, 9I 94.

- MacCulloch R. D., Fu J., Darevsky I. S., Danielyan F., and Murphy R. W. (1995), "Allozyme variation in three closely related species of Caucasian rock lizards (*Lacerta*)," *Amphibia–Reptilia*, **16**, 331 340.
- MacCulloch R. D., Murphy R. W., Fu J., Darevsky I. S., and Danielyan F. (in press), "Disjunct habitats as islands: genetic variability in the Caucasian rock lizard *Lacerta portschinskii*," *Genetica*.
- Mayer W. and Lutz D. (1989), "Chemosystematische Untersuchungen zur Phylogenes der Sammelgattung *Lacerta* (Reptilia: Sauria: Lacertidae)," *Z. Zool. Syst. Evol.-Forsch.*, 27, 338 349.
- Murphy R. W., Darevsky I. S., MacCulloch R. D., Fu J., and Kupriyanova L. A. (1996a), "Evolution of the bisexual species of Caucasian rock lizards: a phylogenetic evaluation of allozyme data," *Russ. J. Herpetol.*, 3(1), 18 31.
- Murphy R. W., Sites J. W., Jr., Buth D. G., and Haufler C. H. (1996b), "Proteins: Isozyme electrophoresis," in: *Molecular Systematics*, 2nd Edition, D. M. Hillis, C. Moritz, and B. K. Mable (eds.), Sinauer Associates, Sunderland (Mass., USA).
- Sattler P. W. and Ries J. S. (1995), "Intraspecific genetic variation among four populations of the Texas horned lizard, *Phrynosoma cornutum*," *J. Herpetol.*, 29, 137 141.
- Swofford D. I. and Selander R. B. (1989), BIOSYS-1: A Computer Program for the Analysis of Allelic Variation in Population Genetics and Biochemical Systematics, Release 1.7, Illinois Natural History Survey, Urbana (USA).
- Wright S. (1978), Evolution and Genetics of Populations. Vol. 4. Variability Within and Among Natural Populations, Univ. Chicago Press, Chicago.