



Contents lists available at ScienceDirect

Molecular Phylogenetics and Evolution

journal homepage: www.elsevier.com/locate/ympev

Common lizards break Dollo's law of irreversibility: Genome-wide phylogenomics support a single origin of viviparity and re-evolution of oviparity

Hans Recknagel^a, Nicholas A. Kamenos^b, Kathryn R. Elmer^{a,*}

^a Institute of Biodiversity, Animal Health and Comparative Medicine, College of Medical, Veterinary & Life Sciences, University of Glasgow, Glasgow G12 8QQ, UK

^b School of Geographical and Earth Sciences, University of Glasgow, Glasgow G12 8QQ, UK

ARTICLE INFO

Keywords:

Squamata
 Lacertidae
 Dollo's law
 Transition
 Viviparity
 Biogeography
 Molecular systematics

ABSTRACT

Dollo's law of irreversibility states that once a complex trait has been lost in evolution, it cannot be regained. It is thought that complex epistatic interactions and developmental constraints impede the re-emergence of such a trait. Oviparous reproduction (egg-laying) requires the formation of an eggshell and represents an example of such a complex trait. In reptiles, viviparity (live-bearing) has evolved repeatedly but it is highly disputed if oviparity can re-evolve. Here, using up to 194,358 SNP loci and 1,334,760 bp of sequence, we reconstruct the phylogeny of viviparous and oviparous lineages of common lizards and infer the evolutionary history of parity modes. Our phylogeny supports six main common lizard lineages that have been previously identified. We find strong statistical support for a topological arrangement that suggests a reversal to oviparity from viviparity. Our topology is consistent with highly differentiated chromosomal configurations between lineages, but disagrees with previous phylogenetic studies in some nodes. While we find high support for a reversal to oviparity, more genomic and developmental data are needed to robustly test this and assess the mechanism by which a reversal might have occurred.

1. Introduction

There are numerous examples for the loss of a complex trait in the animal kingdom throughout evolution. Dollo's law of irreversibility states that once such a complex trait has been lost, it cannot be regained (Gould, 1970). Some exceptions to this rule have been discovered, though it remains a very rare phenomenon in evolution (Collin and Miglietta, 2008; Lynch and Wagner, 2010). Oviparity (egg-laying) is an example for such a complex trait and has been lost on several independent occasions throughout animal evolution (Lee and Shine, 1998; Murphy and Thompson, 2011). While there are more than a hundred independent transitions from oviparity to viviparity (live-bearing) in reptiles (Blackburn, 2006; Sites et al., 2011), only one robust example for the re-evolution of the eggshell is known to date (Lynch and Wagner, 2010). Molecular mechanisms by which reversals in complex traits such as reproductive mode occur are to date unknown.

The common lizard (*Zootoca vivipara*) is the most widespread extant terrestrial reptile species. Its distribution covers nearly the whole of Europe, northern and central Asia and as far as Japan in its easternmost range. Within this distribution, common lizards have adapted to various extreme environments. Arguably the most salient of these adaptations is

the evolution of viviparity, unique within European lizards that are otherwise oviparous. As one of the youngest transitions from oviparity to viviparity known in vertebrates (Pyron and Burbrink, 2014; Surget-Groba et al., 2006), common lizards are an emerging model system for the study of viviparity (Freire et al., 2003; Le Galliard et al., 2003; Murphy and Thompson, 2011). However, not all common lizards are live-bearing: of the six currently recognized common lizard lineages, two are oviparous and four are viviparous (Surget-Groba et al., 2006; Fig. 1). One oviparous lineage is restricted to northern Spain and southwestern France, allopatric to all other common lizard lineages. A second oviparous lineage occurs in the southern part of the Alps. Four viviparous lineages cover the rest of the Eurasian distribution (Mayer et al., 2000; Surget-Groba et al., 2006; Fig. 2).

The phylogenetic relationships within *Zootoca* have not been fully resolved. The evolutionary history of the two different parity modes has been controversial depending on which data were used to interpret the phylogenetic relationships. In a first study using a single mitochondrial gene, both oviparous lineages were found to be sister to all other viviparous lineages, consistent with a single origin of viviparity (Surget-Groba et al., 2001; Fig. 1A). However, subsequent analyses on the karyotype of common lizards resulted in a more complex evolutionary

* Corresponding author.

E-mail address: Kathryn.Elmer@glasgow.ac.uk (K.R. Elmer).

<https://doi.org/10.1016/j.ympev.2018.05.029>

Received 25 November 2017; Received in revised form 12 April 2018; Accepted 22 May 2018

1055-7903/© 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

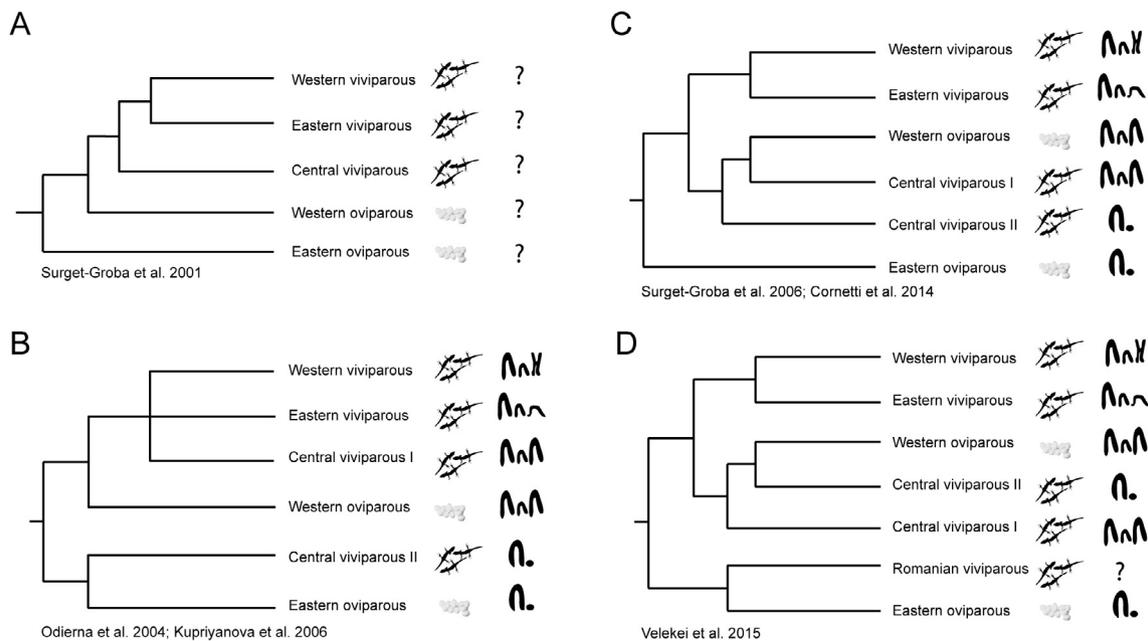


Fig. 1. Alternative hypotheses for phylogenetic relationships of common lizards and parity mode evolution. Parity mode and sex chromosome configuration (ZW or Z_1Z_2W ; Odierna et al., 2004) are illustrated next to each respective lineage. Phylogenetic tree (A) involves a single origin of viviparity and was supported by one mtDNA gene. The second tree (B) is based on karyological studies and suggests two independent origins of viviparity. Hypothesis (C) suggests a reversal to oviparity as most parsimonious scenario, based on mtDNA and a few nuclear genes. The last phylogeny (D) includes a recently discovered viviparous lineage in the Carpathians, which was found to be closely related to the eastern oviparous lineage. Parity mode evolution in this scenario involves two independent origins of viviparity and a reversal to oviparity.

scenario, arguing for two origins of viviparity based on sex-chromosome evolution (Z_1Z_2W or ZW) (Odierna et al., 2004; Surget-Groba et al., 2006; Fig. 1B). More extensive geographic sampling and sequencing of mitochondrial genes instead favored a scenario of a single origin of viviparity followed by a reversal to oviparity in the Spanish western oviparous lineage (Cornetti et al., 2014; Surget-Groba et al., 2006; Fig. 1C), though this phylogeny was incompatible with a single origin of the Z_1Z_2W sex chromosome system. Finally, a population inhabiting the Carpathian region in Romania was discovered recently and was found to be most closely related to the phylogenetically basal eastern oviparous lineage based on mtDNA (Velekei et al., 2015; Fig. 1D). The reproductive mode of this lineage was not reported, but since all other common lizard populations in its geographic proximity are viviparous (Surget-Groba et al., 2006), this would suggest another independent origin of viviparity. However, all phylogenies to date have had limited support at deeper nodes essential for the interpreting the evolutionary scenarios of parity mode evolution. Moreover, phylogenies reconstructed only from mitochondrial DNA have limited information and frequently misrepresent the ‘true’ phylogenetic relationships (Ballard and Whitlock, 2004; Near and Keck, 2013; Wallis et al., 2017). Therefore, it is essential to incorporate high resolution nuclear DNA sequencing to resolve difficult topologies. Moreover, coalescent-based approaches for disentangling incomplete lineage sorting effects and hybridization have considerably advanced phylogenetic reconstruction (Bouckaert et al., 2014; Pickrell and Pritchard, 2012; Posada, 2016).

The evolutionary implications for models involving several origins of viviparity and/or a reversal to oviparity are significant. A reversal to oviparity from viviparity is considered a very unlikely evolutionary scenario, presumably breaking Dollo’s law of irreversibility (Lee and Shine, 1998). Common lizard parity mode evolution could represent one of the very few examples for an exception to this ‘law’ (Surget-Groba et al., 2006). Further, the evolution of both oviparity and viviparity are difficult to study from a molecular genetic perspective because they have most frequently occurred at deep evolutionary time scales. Common lizards provide an example of recent parity mode

changes and therefore a critical insight to usually more ancient evolutionary events.

To tackle this outstanding evolutionary question, we use genome-wide phylogenomics with data from double-digest restriction-site associated DNA sequencing (ddRADSeq), a next generation sequencing (NGS) technique, to identify DNA polymorphisms across all common lizard lineages (Peterson et al., 2012; Recknagel et al., 2015, 2013). Using broad geographic sampling of 67 individuals, we reconstructed a nuclear phylogeny of up to 1.33 million nucleotides, and a mitochondrial DNA phylogeny based on cytochrome *b*, using coalescent, Maximum Likelihood, and Maximum Parsimony methods. We performed topological tests and model-based ancestral state reconstructions to assess the likelihood of alternative scenarios for parity mode evolution. Our results strongly support a single origin of viviparity in common lizards and a subsequent reversal to oviparity in one derived lineage as the most probable scenario of reproductive mode evolution.

2. Material and methods

2.1. Sampling

Samples and specimens were obtained from the Natural History Museum in Vienna, the Royal Ontario Museum, and fieldwork during 2013–2016 (see Table S1 for specimens and Fig. 2 for a map of collecting localities). Lizards were collected by diurnal opportunistic searches. Tail clips (up to 2 cm) were extracted and preserved in 95–99% ethanol and lizards were released thereafter. Mode of reproduction was assessed by observation of an individual retained in captivity until oviposition/parturition or from data on other individuals at the same site.

2.2. Generation of molecular data

DNA was extracted from tissue using a Dneasy Blood and Tissue Kit (Qiagen) following the manufacturer’s protocol. Three genomic libraries were constructed using double-digest restriction-site associated

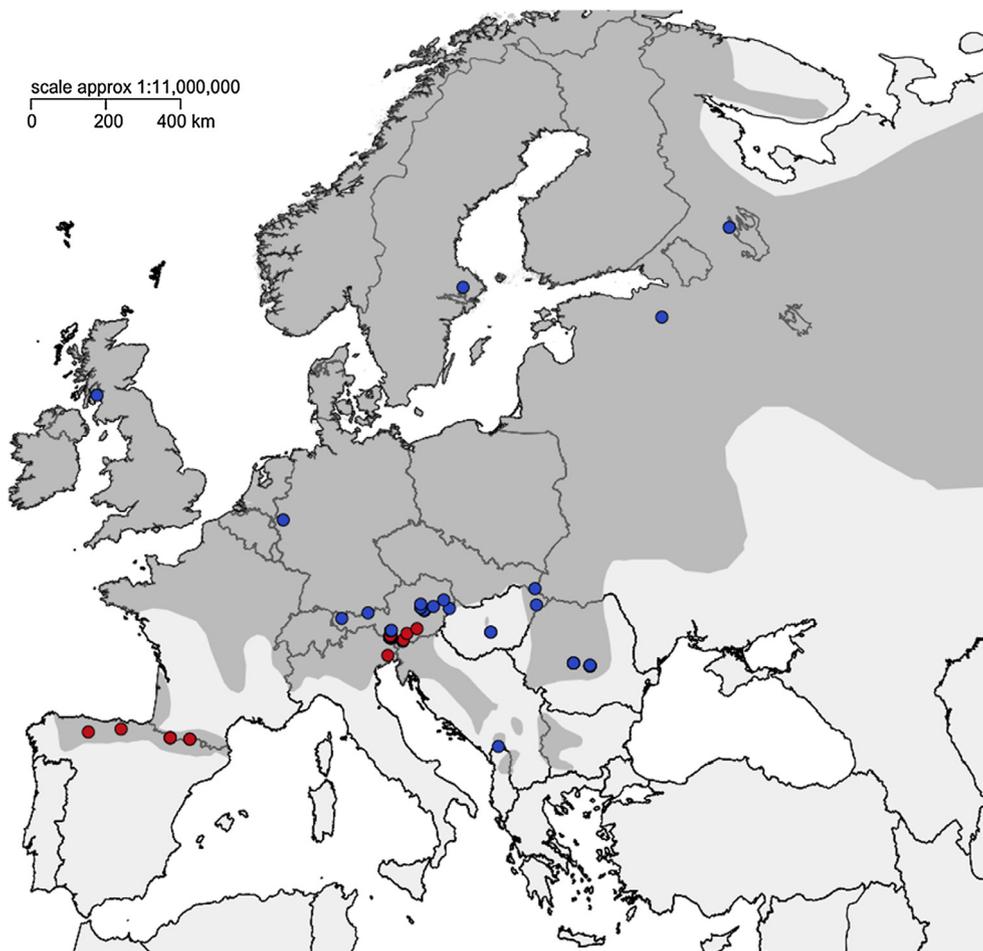


Fig. 2. Map of common lizard (*Zootoca vivipara*) sampling locations within Europe. The dark grey shaded area marks the distribution of the common lizard in Europe. Each dot represents a single individual (red = oviparous; blue = viviparous) captured at the respective location. Note that a single individual from central Russia included in the phylogenetic analyses is outside the scope of the map (see Table S1). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

DNA sequencing (ddRADSeq). The first two libraries were run on an IonProton sequencing machine with a median of 96 bp read length (ddRADSeq-ion; Recknagel et al., 2015) and the third library was paired-end sequenced on an Illumina HiSeq 4000 with 150 bp read length. Briefly, 1 ug of starting DNA material was digested using restriction enzymes *Pst*I-HF and *Msp*I and subsequently cleaned with the Enzyme Reaction Cleanup kit (Qiagen). Following purification, the amount of DNA in each individual was normalized to the sample with the lowest concentration within a library (237 ng in first, 400 ng in second, and 275 ng in third library) to minimize coverage variation. Platform specific barcoded (for IonProton: A-adapter, for Illumina: P1 adapter; binding to *Pst*I-HF overhang) and global (for IonProton: P1-adapter, for Illumina: P2 adapter; binding to *Msp*I overhang) adapters were ligated to the sticky ends generated by restriction enzymes. The ligated DNA fragments were then multiplexed and size-selected using a Pippin Prep (Sage Science) for a range between 175 and 225 bp for the IonProton platform and 150–210 bp for Illumina. To assure that the same set of loci are selected between platforms, size selection ranges were adjusted because adapter lengths are not the same between platforms. Seven separate PCR reactions (for details see Recknagel et al., 2015) were performed per genomic library and combined (Peterson et al., 2012). Following PCR purification, libraries were electrophoresed on a 1.25% agarose gel to remove any remaining adapter dimers and fragments outside the size range selected by the Pippin Prep. SYBRsafe (Life Technologies) was used for gel staining and bands in the size selected range were cut out manually and DNA was extracted

from the matrix using a MinElute Gel Extraction Kit (Qiagen). Following the gel extraction, DNA was quantified using a Qubit Fluorometer with the dsDNA BR Assay. Quality and quantity of genomic libraries was assessed using a TapeStation or Bioanalyzer (Agilent Technologies). The first two libraries were sequenced at Glasgow Polyomics using an Ion PI Sequencing 200 Kit v3 on an Ion Proton PI chip at a target read size of 100 bp. The third library was sequenced at Edinburgh Genomics on an Illumina HiSeq 4000 machine with paired-end sequencing of 150 bp reads.

In addition to ddRADseq, mitochondrial DNA (mtDNA) from cytochrome *b* with primers MVZ04H and MVZ05L (~430 bp) was amplified (Smith and Patton, 1991) and PCR products were sequenced with the forward primer (MVZ04H) on an ABI 3130x at Dundee University. Sequences were quality checked by eye, and trimmed and aligned using Geneious v. 7.1.9 (Kearse et al., 2012). Data are deposited in NCBI (Genbank accessions MH395870-MH395927; BioProject PRJNA472940).

2.3. Bioinformatic analysis

All NGS generated reads were analyzed using the RADseq software tool STACKS v.1.41 (Catchen et al., 2011). Libraries were demultiplexed and reads were trimmed to a common length of 70 bp to maximize the number and length of retained reads (Recknagel et al., 2015). Each individual was then aligned to a *Zootoca vivipara* reference genome v. 0.9 (Yurchenko et al. in prep) using bwa (Li and Durbin,

2010) and samtools (Li et al., 2009). A catalogue of all loci identified across individuals was subsequently created using the genome referenced stacks from each individual with a minimum coverage of 3x per individual locus.

Missing data can have a substantial impact on phylogenetic inference from NGS generated data and can vary between taxonomic and phylogenetic levels (Eaton et al., 2017; Jiang et al., 2014; Rowe et al., 2011; Streicher et al., 2016). Therefore, it is crucial to first evaluate the impact of missing data before phylogenetic analysis. We filtered our data with two main options: (i) using a variable minimum number of individuals that a locus had to be present in, and (ii) varying the number of SNPs per locus from one to three. The amount of missing data was increased from 0% to 90% at 10% intervals. For each of these categories, loci containing only a single SNP, two SNPs, three SNPs and one to three SNPs were extracted from the whole dataset. These datasets were extracted to test the impact of missing data and number of SNPs on phylogenetic resolution and to assess optimal settings for data extraction.

2.4. Phylogenetic analysis

Suitability of data sets that differed in degree of missing data and number and type of SNP loci was assessed by comparing the sum of bootstrap supports (at deep, at shallow, and at all nodes combined) (Huang and Lacey Knowles, 2016). The best performing dataset for inferring the evolutionary history of parity mode in common lizards was identified and chosen for more exhaustive phylogenetic and comparative analyses. This best performing dataset was assessed by constructing Maximum-likelihood (ML) phylogenies using the software RAxML vers. 8.1.20 with a GTRGAMMA substitution model of evolution (Stamatakis, 2006). Conditions producing the highest bootstrap sum phylogeny were the ones chosen for all subsequent analyses.

We inferred Maximum-likelihood (ML) phylogenies using RAxML. ML bootstrap search with 100 replicates using a GTRGAMMA model was performed. Support values were applied to the best scoring ML tree. An initial phylogenetic analysis including the outgroup species *Iberolacerta horvathi* identified the eastern oviparous clade as sister to all five other *Zootoca* lineages with high confidence (bootstrap support 100), as has been shown by previous analyses (Cornetti et al., 2014; Mayer et al., 2000; Surget-Groba et al., 2006). We further used ADMIXTURE (vers. 1.3.0; Alexander et al., 2009) to test for genetic coherence of the main *Zootoca* lineages. ADMIXTURE assesses the genomic ancestry of individuals according to a given set of genetic clusters. A variable number of genetic clusters k was run, from 1 to 6 k and best fit inferred from ten-fold cross-validation. The genetic cluster with the lowest cross-validation error was chosen as optimal k . These analyses suggested monophyly of the six main lineages and limited levels of admixture. Pairwise genetic differentiation between lineages was assessed using the R package *diveRsity* (Keenan et al., 2013).

((((The topology of the best ML tree was compared to alternative topologies that had been proposed in previous studies. These topologies included scenarios consistent with a single origins of viviparity (Mayer et al., 2000; Surget-Groba et al., 2001), multiple origins of viviparity (Odierna et al., 2004) and a single evolution of viviparity with a reversal to oviparity (Surget-Groba et al., 2006; Velekei et al., 2015) (Table 1). We computed per site log likelihoods for each of the five trees and used these to perform Approximately Unbiased tests (AU tests) (Shimodaira, 2002), Shimodaira-Hasegawa tests (SH tests) (Shimodaira and Hasegawa, 1999), Kishino-Hasegawa tests (KH tests), and Bayesian posterior probabilities (PPs) calculated by the BIC approximation as all implemented in CONSEL vs. 0.1a (Shimodaira and Hasegawa, 2001).

We performed a Bayesian approach to infer the topology in BEAST2 (Bouckaert et al., 2014). For this approach, we included a full alignment of all RAD loci (19,068 RAD loci; 1,334,760 total bp; 84,017 variant sites). The number of total SNPs differs from other analyses as loci were set to be present in at least 40% of individuals of each of the

six lineages, instead of just being present in at least 40% of individuals across the whole phylogeny. We used the GTRGAMMA substitution model. The analysis was run on CIPRES (Miller et al., 2010) for 500 million generations, sampling trees every 50,000 and discarding 10% as burn-in. Convergence was assessed in TRACER (Rambaut and Drummond, 2009) and accepted if ESS values of all parameters were larger than 100. In addition, we reconstructed the species tree using ASTRAL (Mirarab et al., 2014), which is based on the multi-species coalescent model. The software package reconstructs evolutionary relationships between species (or deep lineages) with an algorithm integrating over all possible gene trees. Monophyletic lineages were identified from the previous Maximum likelihood analyses as highly supported (BS = 100) evolutionary deep clusters of individuals. Each clade contained a minimum number of nine individuals (ranging from 9 to 16 individuals). *Iberolacerta horvathi* was included as an outgroup species. ML gene trees from 375,103 RAD loci were reconstructed in RAxML under the GTRGAMMA substitution model using a window size of 100 sites. This resulted in 3537 gene trees that were used as an input file in ASTRAL.

Additional phylogenetic analyses were carried out under the Maximum Parsimony (MP) optimality criteria. We performed a heuristic bootstrap search with 2000 replicates carried out in PAUP* (Swofford, 2002) using TBR branch swapping and with ten random addition sequence replicates for each bootstrap replicate. The 50% consensus bootstrap tree was compared to phylogenies generated with ML and Bayesian analyses.

To incorporate potential past migration events and incomplete lineage sorting effects, we performed a TREEMIX v.1.3 (Pickrell and Pritchard, 2012) search using only independent SNPs (one SNP per locus; 49,107 loci included) and a window size of 1000 bp. We included zero to six migration events and compared the variance explained between resulting trees with and without migration events to evaluate the impact of migration. We calculated f_3 -statistics to assess whether admixture has played a role in the evolution of common lizard lineages.

For the mitochondrial dataset, we performed a bootstrap ML search using RAxML (100 bootstrap replicates), MP using the same parameters mentioned above and Bayesian reconstruction with BEAST2 to generate the phylogeny. The best substitution model for BEAST2 was inferred from eleven different substitution schemes in JMODELTEST2 (Darriba et al., 2012) based on lowest AICc and run on CIPRES. We ran BEAST2 for 20 million generations and discarded 10% as burn-in. Convergence was inferred if ESS values in TRACER were larger than 100.

We tested different models of ancestral trait reconstruction to assess the likelihood of a reversal to oviparity using different transition rates from oviparity to viviparity and viviparity to oviparity (Goldberg and Igić, 2008). We used the corHMM package in R (Beaulieu et al., 2012) which reconstructs ancestral states of binary characters allowing transition rates to differ and treating them as hidden states in a Markov process. We used the ML tree retrieved from RAxML as the input tree for the ancestral trait reconstruction using marginal likelihoods within the rayDisc function. With the exception of the derived viviparous species *Eremias multiocellata* from Asia, common lizards are the only viviparous taxon within the family of Lacertidae (Pavlicev and Mayer, 2009). At least the last five deeper ancestral nodes were all oviparous (Pyron et al., 2013). Therefore, the root state for common lizards was fixed to oviparity. First, we tested which evolutionary scenario was favored under an ‘all rates different’ model of evolution. Second, a model with no transitions from viviparity to oviparity, conferring to Dollo’s law, was applied. Third, we used a published transition rate from a phylogeny across squamates (Pyron and Burbrink, 2014). Finally, using rates from the first model as initial values, we gradually decreased the transition rate from viviparity to oviparity to test at which point the likelihood of a ‘no reversal’ scenario was i) equally likely and ii) 90% more likely than a scenario including a reversal to oviparity. Models were compared using AICc values and likelihood ratio tests.

3. Results

3.1. Data evaluation and identification of optimal parameters for phylogenomic datasets

The total number of generated reads was 828,000,972 (1st library: 10,000,000 reads, 2nd library: 42,377,658 reads, 3rd library: 775,623,314 paired-end reads). After sorting reads into individual loci, mean coverage per individual was 27.6x with a standard deviation of 11.0x (range: 9.2x–66.9x; median: 24.1x).

We found that phylogenetic resolution generally improved by accepting larger numbers of individuals with missing data (Fig. S1). The best summed bootstrap support was achieved using loci that were present in at least 40% of all individuals. Accepting more missing data did not improve phylogenetic resolution. The highest number of SNPs (including up to three SNPs) resulted in the overall highest phylogenetic resolution (Fig. S1). Therefore, we chose the dataset with loci present in at least 40% of all individuals and including all SNPs (no restriction on number of SNPs per locus) for all subsequent analyses. Genotyping error was low (2.0–2.9% per SNP) based on three technical replicates and comparable to previous studies (Mastretta-Yanes et al., 2015; Recknagel et al., 2015).

3.2. Mitochondrial DNA phylogeny

The final alignment of the cytochrome *b* gene consisted of 428 bp (42 parsimony informative sites). HKY + I was identified as the best substitution model for BEAST2 (Table S2). This phylogeny resolved eastern oviparous, central viviparous, and western oviparous each as monophyletic (Fig. S2). However eastern viviparous, central viviparous, and western viviparous lineages were all polyphyletic, suggesting considerable introgression and a poor association of single gene mtDNA with the phylogeny generated from genome-wide data. Support values were generally considerably lower for both deeper and terminal nodes compared to the phylogeny generated from the extensive genomic dataset. The topology also differed considerably from the topology generated from phylogenomic data (Fig. 3; Fig. S2).

3.3. Monophyletic clades in *Zootoca vivipara* and reconstruction of evolutionary history

All phylogenomic reconstructions returned six monophyletic evolutionary divergent lineages with high confidence (all MP and ML bootstrap supports of 100 and PP of 1.0; Fig. 3). The eastern oviparous lineage was sister to all other lineages, followed by central viviparous II. The remaining four lineages were split into two groups; one with the western oviparous and central viviparous I lineages and the other with the eastern and western viviparous lineages. This topology is concordant with a single origin of viviparity and a reversal to oviparity in the western oviparous lineage (see 3.4 for topological analyses). Population structure also confirmed these six genetic lineages, with high average membership values for each respective lineage (mean Q-values ranged from 92 to 100% identity within each lineages) (Fig. 3). These six lineages correspond to phylogeographic clades that were previously identified. The recently reported distinct Carpathian haplotype (Velekei et al., 2015) was not confirmed as a separate genetic cluster in our phylogenomic reconstruction and was nested within the eastern viviparous lineage (individuals ELT07086–ELT07095). Our mitochondrial dataset confirmed monophyly of some of the lineages with good support (eastern oviparous, central viviparous I, western oviparous), while others were not supported (Fig. S2). In contrast to the nuclear data, the separate Carpathian clade was strongly confirmed by mitochondrial DNA as monophyletic and sister to the eastern oviparous lineage (Fig. S2).

Genetic differentiation between all six lineages was substantial (Table S3). *Fst* and *Jost's D* values were largest between eastern

oviparous and all other lineages (*Fst*: 0.42–0.52; *Jost's D*: 0.013–0.018), and second largest between western oviparous and all other lineages (*Fst*: 0.35–0.51; *Jost's D*: 0.007–0.016), indicating that these are highly differentiated lineages. Compared to *Fst*, *Jost's D* was weaker between the western oviparous and all other viviparous lineages (Table S3). Genetic differentiation between the viviparous lineages was less pronounced (*Fst*: 0.23–0.32; *Jost's D*: 0.004–0.008).

3.4. Evolutionary scenarios for parity evolution

We found significant support for topologies associated with a single origin of viviparity and a reversal to oviparity. Bayesian, Maximum likelihood and parsimony analyses all confirmed the same topological configuration for the six main common lizard lineages with high nodal supports (bootstraps = 100, all posterior probabilities = 1.0) (Fig. 3). Phylogenies from all reconstruction methods support a topology in which the eastern oviparous lineage is sister to all other lineages. The following lineage splitting is the central viviparous II lineage, sister to all remaining lineages. The western oviparous lineage is nested within the viviparous lineages, sister to the central viviparous I lineage. This topology suggests a single origin of viviparity in common lizards and a reversal to oviparity in the western oviparous lineage as the most parsimonious scenario for parity mode evolution. The species tree reconstruction also supported a single origin of viviparity and a reversal to oviparity as the most parsimonious evolutionary scenario (Fig. S3). The topology at deeper nodes was similar to the relationships recovered with other reconstruction methods, but differed in the relationship between the western oviparous, central viviparous I, western viviparous and eastern viviparous (Fig. S3). In the species tree, the western oviparous lineage was sister to the eastern viviparous lineage, and the central viviparous I was sister to the western viviparous lineage. All nodes had high posterior probabilities (> 0.98).

Using monophyly constraints and statistical topology testing, alternative scenarios of parity mode evolution were unlikely. Alternative scenarios included: a single origin of viviparity, multiple independent origins of viviparity, a reversal to oviparity but independent sex chromosome evolution, and multiple origins of viviparity and a reversal to oviparity (Table 1), all of which were less likely than a single origin of viviparity, a reversal to oviparity, and a single change in sex chromosome configuration, i.e. a topology consistent with Fig. 3.

Ancestral trait reconstructions also confirmed that a single origin of viviparity followed by a reversal to oviparity in the western oviparous lineage was the most likely scenario. A model with no reversal to oviparity, consistent with Dollo's law of irreversibility, was significantly less likely ($\Delta\text{AICc} = 11.18$, $P < 0.001$; Table 2). After decreasing the transition rate from viviparity to oviparity to 0.0039, or 180-fold, ancestral states for internal nodes on which a transition occurred had equal probabilities of being viviparous or oviparous, and after decreasing it to 0.0004, or 1600-fold, ancestral states had a likelihood of 90% being oviparous. Both models performed significantly worse than the model including a reversal to oviparity (equal probability: $\Delta\text{AICc} = 7.05$, $P < 0.01$; 90% probability oviparous: $\Delta\text{AICc} = 8.23$; $P < 0.001$; Table 2). For a scenario of multiple independent origins of viviparity and no reversal to oviparity, the transition ratio from viviparity to oviparity had to be 4300x smaller than the rate from oviparity to viviparity to result in 90% likelihood of oviparous ancestors (Table 2).

Reconstructing evolutionary relationships between the six main phylogenetic lineages in TREEMIX results in a similar topology as retrieved from the other analyses, with eastern oviparous consistently sister to all other lineages. The overall likelihood and variance explained by the tree increased when including more migration events, and reached a plateau after two migration events (Fig. S4). Topologies were unstable when more migration events were included, though these topological changes should be considered with caution since all f_3 -statistics were positive, indicating that admixture has not played a

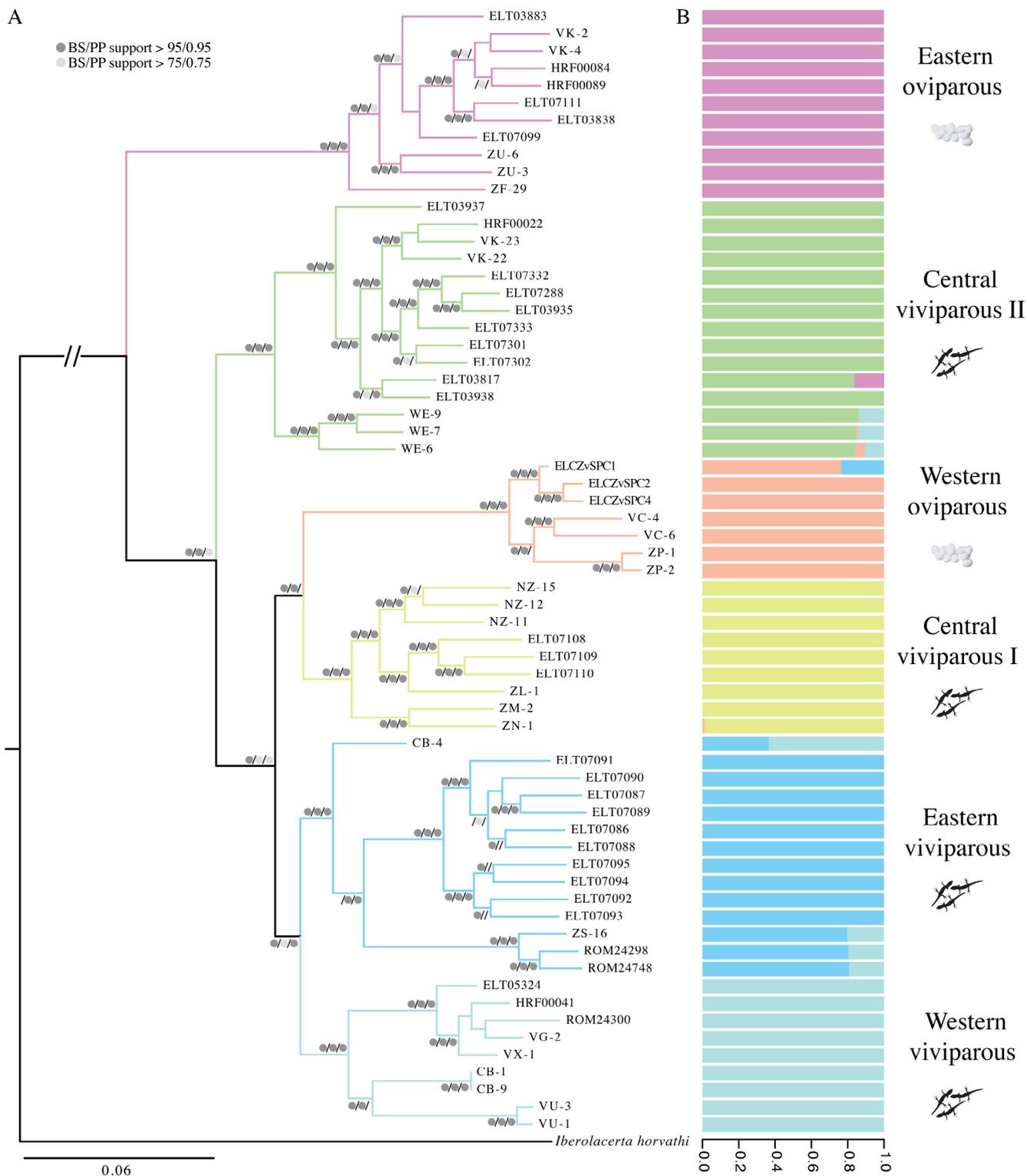


Fig. 3. Bayesian (B), Maximum likelihood (ML) and maximum parsimony (MP) reconstruction of common lizard evolutionary relationships based on ddRADSeq data. (A) The Bayesian tree was used with a full alignment using 1,334,760 sites (84,017 SNPs) and ML and MP trees were constructed with 194,358 SNPs. (B) An ADMIXTURE analysis included the 194,358 SNPs and a k of 6 genetic clusters. Individuals are aligned vertically and respective membership values for each genetic cluster are illustrated. Parity mode and lineage are indicated on the right. *Iberolacerta horvathi* was used as an outgroup (true branch length not shown for graphical reasons).

Table 1

Statistics of alternative topological constraints. Five alternative topological constraints were compared to the best performing maximum likelihood tree. Topological constraints were set to represent different evolutionary hypotheses of parity mode evolution (assuming the most parsimonious path of evolution, i.e. the lowest number of possible transitions). Constraint models are ranked by observations, starting with the model without constraint. Constraint models are the following: (i) ‘no constraint’ is consistent with a reversal to oviparity and refers to the topology in Fig. 3, (ii) ‘viviparous CVII basal’ is the same topology as (i) but specifying that the eastern oviparous lineage is an outgroup and the central viviparous II lineage is sister to all remaining lineages; it is consistent with a reversal to oviparity and Fig. 3, (iii) ‘multiple viviparity’ constrains central viviparous II as sister to eastern oviparous, and western oviparous sister to all other viviparous lineages, consistent with two independent origins of viviparity and Fig. 1B, (iv) ‘oviparity basal’ constrains all viviparous lineages to an ingroup sister to the oviparous lineages and is consistent with a single origin of viviparity and Fig. 1A, (v) ‘viviparous CVII not basal’ constrains the eastern oviparous lineage to be sister to all other lineages, but does not constrain CVII to be sister to all other viviparous lineages; it is consistent with a reversal to oviparity but not with sex chromosome evolution and corresponds to Fig. 1C, and (vi) ‘viviparous RO with EO’ constrains the Carpathian lineage to be sister to the eastern oviparous lineage, consistent with multiple independent origins of viviparity and potentially a reversal to oviparity and corresponds to Fig. 1D.

Constraint	Rank	obs	AU	NP	BP	PP	KH	SH	wtd-KH	wtd-SH
No constraint	1	0	0.518	0.493	0.502	0.500	0.496	0.918	0.496	0.918
Viviparous CVII basal	2	0	0.535	0.501	0.494	0.500	0.504	0.891	0.504	0.891
Multiple viviparity	3	404.6	0.000	0.000	0.000	0.000	0.000	0.021	0.000	0.000
Oviparity basal	4	452.7	0.005	0.004	0.004	0.000	0.004	0.004	0.004	0.011
Viviparous CVII not basal	5	1206.9	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
Viviparous RO with EO	6	2478	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000

Abbreviations are: obs = observations, AU = Approximately unbiased test, NP = non-scaled bootstrap probability, BP = bootstrap probability, PP = Bayesian posterior probability, KH = Kishino-Hasegawa test, SH = Shimodaira-Hasegawa test, wtd = weighted, CVII = central viviparous II, CVI = central viviparous I, RO = Carpathian viviparous clade, EO = eastern oviparous.

Table 2

Models of ancestral trait reconstruction. For all models, root state was fixed to oviparity. Transition rates (q_{01} = oviparity to viviparity; q_{10} = viviparity to oviparity) were manipulated to test the likelihood of models including no reversal from viviparity. The second model with no reversal is consistent with Dollo’s law of irreversibility. In the third model, published transition rates across the squamate tree were used. In the last two models, transition rates were altered until ancestral states for internal nodes on which transitions occurred had a likelihood of (i) 50% oviparity and viviparity and (ii) 90% oviparity.

Model	q_{01}	q_{10}	q_{01}/q_{10}	$-\ln L$	AIC	AICc	$\Delta AICc$
Null model	1.892	0.7090	3	-8.02	20.05	20.24	0
No reversal	1	0	NA	-13.61	31.23	31.42	11.18
Across squamates	0.05	0.006	8	-14.40	32.90	33.10	12.86
Equal prob.	1.892	0.0039	480	-11.55	27.10	27.29	7.05
No reversal more likely	1.892	0.0004	4300	-12.14	28.28	28.47	8.23

major role in the evolution of common lizard lineages (Table S4).

4. Discussion

4.1. Evolutionary history of parity mode evolution

Here we show that the most parsimonious scenario for the evolution of parity mode in common lizards includes a single origin of viviparity and a reversal to oviparity in one lineage (western oviparous). While other lacertid relatives and the outgroup common lizard lineage (eastern oviparous) are oviparous, all other common lizard lineages are viviparous, except for the western oviparous lineage nested within a viviparous clade (Fig. 3). Our genome-level phylogeny based on up to 1,334,760 nucleotides was highly supported by Bayesian, ML, and MP analyses (support values > 0.95). Topologies compatible with other parity mode scenarios, such as no reversal to oviparity or multiple origins of viviparity (per Fig. 1A, B, D) performed significantly worse in all statistical tests (Table 1). A scenario of three independent origins of viviparity from oviparity, alternatively compatible with our topology, would only be likely if the transition rate to oviparity is extremely low (Table 2). The species tree reconstruction supported the same evolutionary scenario compatible with a single origin of viviparity and a reversal to oviparity in the western oviparous lineage. Ancestral trait reconstructions also found that models not allowing a reversal from viviparity to oviparity were significantly less likely compared to the

optimal model including a reversal to oviparity. We find considerable differences between our high resolution phylogenomic tree and our mtDNA phylogeny (Fig. S2).

The evolution of oviparity and viviparity in common lizards has been contentious and a range of studies, using different geographic and genetic sampling, have failed to converge on an evolutionary scenario. To date, mitochondrial DNA, nuclear DNA, and karyotypic markers have not agreed on a single topology (Fig. 1; Odierna et al., 2004; Surget-Groba et al., 2006, 2001; Velekei et al., 2015). For example, previous research suggested that a reversal to oviparity occurred in common lizards, however support was based on only limited data and support (Cornetti et al., 2014; Surget-Groba et al., 2006). It has also been proposed that viviparity evolved multiple times independently (Odierna et al., 2004; Velekei et al., 2015), however, these studies were limited to the use of a single marker. Our phylogeny is the first that is consistent with nuclear genetic markers and chromosomal configuration (Fig. 1; Fig. 3).

Other aspects of common lizard genetics and reproductive traits also support our inference of a reversal to oviparity. The eastern oviparous and western oviparous lineages have different morphological and physiological reproductive characteristics, such as thinner eggshells and shorter incubation time (Arrayago et al., 1996; Lindtke et al., 2010). We suggest this is compatible with our phylogeny; the derived oviparous lineage is due to a reversal to oviparity instead of retaining the ancestral oviparous condition, and in doing so the thickness of the eggshell is reduced. Our phylogeny is consistent with the most parsimonious scenario for the derived chromosomal features in common lizards. While both the eastern oviparous and central viviparous II lineages have 36 chromosomes and a ZW sex chromosome configuration, all other lineages exhibit 35 chromosomes and a Z_1Z_2W sex chromosome configuration (Kupriyanova et al., 2008; Odierna et al., 2004; Fig. 1). Previous genetic studies were inconsistent with this derived sex chromosome configuration by placing central viviparous II nested within lineages exhibiting the Z_1Z_2W chromosome configuration instead of being sister to lineages with the derived configuration (Cornetti et al., 2014; Surget-Groba et al., 2001, 2006). The phylogeny presented here is the first molecular phylogeny consistent with a single transition in sex chromosome configuration, changing from the ancestral ZW system to the derived Z_1Z_2W system (Kupriyanova et al., 2006; Odierna et al., 2004).

Calcified eggshell and the associated reproductive life history traits of oviparity represent a complex character that once lost is unlikely to re-evolve, making it a trait long regarded to be subjected to Dollo’s law

of irreversibility (Gould, 1970; Lee and Shine, 1998; Shine and Lee, 1999; Sites et al., 2011). However, research on the re-evolution of insect wings (Collin and Miglietta, 2008; Whiting et al., 2003), snail coiling (Collin and Cipriani, 2003), or mandibular teeth in frogs (Wiens, 2011) has shown that in some cases complex characters can indeed re-evolve. In squamate reptiles, one example exists arguing for the re-evolution of oviparity in sand boas (Lynch and Wagner, 2010). In this example, a scenario with no reversal to oviparity required three additional evolutionary transitions compared to the most parsimonious scenario with a single reversal to oviparity. In addition to the support from parsimonious trait reconstruction from the phylogeny, sand boas lack the egg tooth, which is an important anatomical structure for hatching from eggs that is present in related oviparous snake species. This provides independent evidence for the derived state in sand boas and the re-evolution of oviparity (Lynch and Wagner, 2010). In general, in addition to support from phylogenetic reconstruction, it should be best practice to assess whether the trait re-evolved is developmentally and anatomically similar to the ancestral trait. Substantially different features of the trait in the derived compared to ancestral form can be considered additional evidence for re-evolution, rather than the less plausible scenario that the ancestral form was retained but changed over time while an alternative trait was independently lost in multiple related lineages. In common lizards, the short timespan between the origin of viviparity and the re-evolution of oviparity might have facilitated the reversal, in that not many genomic changes were required. In general, a trait as complex as viviparity is thought to require several changes in the genome (Murphy and Thompson, 2011). Alternatively, it is conceivable that oviparity was reacquired by adaptive introgression from a common lizard lineage exhibiting oviparity. Importantly, this can be tested in the future using a combination of whole genome sequencing techniques, population genomics, and understanding the genetic basis of the trait (Racimo et al., 2015; Stern, 2013).

Whether reversals to oviparity from viviparity occurred frequently in squamate reptiles remains a highly controversial topic. Erroneous phylogenetic reconstruction and limited assessment of characteristics of the trait in question have led to the publication of controversial examples of re-evolution (e.g. Fairbairn et al., 1998; Pyron and Burbrink, 2014) that have been criticized heavily (Blackburn, 2015, 1999; Griffith et al., 2015; King and Lee, 2015; Shine and Lee, 1999; Wright et al., 2015). Moreover, incomplete lineage sorting and/or introgression of the trait in question, combined with the limited molecular information included in most phylogenetic reconstructions, can lead to wrong conclusions in trait evolution (Hahn and Nakhleh, 2016). While here we found substantial support for the re-evolution of oviparity based on the largest genomic dataset to date, more knowledge on the development and genetics of the trait is necessary to unequivocally assess whether a reversal to oviparity occurred in common lizards. In the future, more refined reconstructions using whole genome and phylogenomic data combined with insights into the genetic mechanisms involved in parity mode evolution should provide answers on whether reversals to oviparity occur in squamates and how common they are.

4.2. Evolutionary relationships between common lizard lineages and comments on taxonomic status

Our genome-wide phylogeny recovered a new topology, but this included similar clades as previously supported by mitochondrial DNA reconstructions, except for the Carpathian clade, which we find is nested within the eastern viviparous lineage (Fig. 1; Fig. 3; Fig. S3). Incongruence between nuclear data and mitochondrial data is observed frequently (Ballard and Whitlock, 2004; Near and Keck, 2013; Rodríguez et al., 2017; Wallis et al., 2017). Consistent with previous phylogenetic analyses (Cornetti et al., 2014; Surget-Groba et al., 2006, 2001), we found the eastern oviparous lineage is sister to all other common lizard lineages. Splitting order for the other lineages differs from previous phylogenetic reconstructions, however, the reciprocal

monophyly of all remaining five lineages was highly supported by all analyses here. In agreement with this, f_3 -statistics suggest that there was no significant admixture between lineages (Table S3). Past mitochondrial DNA introgression and capture are a possible mechanism explaining the discordance between mitochondrial and nuclear genes (Leavitt et al., 2017; Willis et al., 2014).

Based on the strong reciprocal monophyly of the lineages, we suggest that a future revision of the subspecific taxonomy may be warranted. Some have argued that *Z. v. carniolica* should be recognized as a separate species based on limited gene flow and reproductive isolation (Cornetti et al., 2015a, 2015b). However, while hybridization is rare and might be geographically restricted, it does occur between *Z. v. carniolica* and other viviparous common lizards (Lindtke et al., 2010; pers. obs.) and phenotypic differences are generally small (Guillaume et al., 2006; Rodríguez-Prieto et al., 2017). Currently, only *Z. v. louislantzi* (western oviparous) is recognized as a subspecies, while other lineages have inconsistent subspecific designations (Arribas, 2009; Schmidler and Böhme, 2011). While diagnostic morphological features are scarce (Guillaume et al., 2006), in-depth analyses using more levels of the phenotype (e.g. differences in colouration, behavior, reproduction and ecology) should resolve whether the distinguished genetic lineages are supported by phenotypic data. A more balanced genetic sampling across the whole geographic range using modern molecular and phylogenetic techniques combined with morphological and ecological data collection of the group is much needed.

4.3. Advantages and challenges of RADSeq data for phylogenetic reconstruction

Our phylogenetic reconstruction represents the most comprehensive and robust phylogeny of common lizards to date, based on 194,358 polymorphic SNPs and 67 individuals. Previous phylogenetic studies on common lizards using mitochondrial data (Surget-Groba et al., 2006) or fewer nuclear markers (Cornetti et al., 2014) had only moderate congruency between different markers and weak support at deeper nodes. In agreement with the challenges from previous studies, our mtDNA phylogeny of an established, informative locus was not compatible with our phylogenomic dataset, highlighting the limitations of mtDNA (Ballard and Whitlock, 2004; Wallis et al., 2017; Willis et al., 2014) and suggesting it is not an appropriate marker for resolving the history of common lizards. More generally, we suggest that for groups with short internal branches and evolutionary histories of recent to several million years divergence, the type of data produced by RADSeq might be optimal to resolve difficult evolutionary splits. This is the case for adaptive radiations or more generally for short and quick speciation events and complex phylogeographic histories (Giarla and Esselstyn, 2015; Rodríguez et al., 2017). This study evidences the power of fast evolving loci (loci with several SNPs) to resolve short phylogenetic branches.

A challenge of short-read phylogenomics and loci with multiple SNPs is the validity of orthology between loci. We show that topological groupings are more robustly supported when using loci with multiple SNPs (Fig. S1) and we present an assessment pipeline for validating the cut-offs for missing data and SNPs per locus. Without a reference genome and a large amount of duplicated and/or repetitive DNA, the orthology of RAD loci is usually not evaluated. Using a reference genome to map the RAD loci and high sequencing coverage per individual, such as done here, are important methodological considerations to overcome these issues (Mastretta-Yanes et al., 2015; Shafer et al., 2017). Disadvantages of these large but informative datasets are long computational time for some analyses, in particular phylogenetic reconstructions using Bayesian coalescence based analyses (Bryant et al., 2012). Advances in phylogenomic methodologies to accommodate these more complex datasets will be important for advancing the field (Delsuc et al., 2005; Fuentes-Pardo and Ruzzante, 2017; Leavitt et al., 2016).

4.4. Conclusions

Our results support a single origin of viviparity in common lizards and a subsequent reversal to oviparity in one derived lineage as the most probable scenario of reproductive mode evolution (Fig. 3, Table 1). In the light of karyological and reproductive data (Arrayago et al., 1996; Heulin et al., 2002; Lindtke et al., 2010; Odierna et al., 2004, 1998), these findings are strong support that a reversal to oviparity has occurred in what is now the allopatric western oviparous lineage (Figs. 2 and 3). More generally, this suggests that Dollo's law of irreversibility is not without exceptions, and might be particularly prone to switches between characters at early stages of trait evolution. For the future, we suggest that common lizards represent an ideal candidate to investigate the genomic basis for evolutionary complex reversals.

Acknowledgments

This work would not have been possible without the invaluable support and contribution of samples by the late Werner Mayer, to whom we are very grateful. We thank B. Murphy and A. Lathrop at the Royal Ontario Museum for providing tissue samples. We particularly thank Austrian and Scottish authorities for issuing collection permits (HE3-NS-959/2013; SNH license number 64972). We thank Megan Layton, Henrique Leitão, Mark Sutherland, Ruth Carey, Michael Andrews, Jade McClelland, and Nathalie Feiner for assistance and companionship in the field during the collection of crucial samples. We thank Aileen Adams, Arne Jacobs, Julie Galbraith, Jing Wang, Lorraine Glennie, and Peter Jeffrey Koene for their help in the lab and Andrey Yurchenko for access to the reference genome and valuable discussions. For funding we gratefully acknowledge a Heredity Fieldwork grant by the Genetic Society to HR, a University of Glasgow Lord Kelvin-Adam Smith PhD Studentship to KRE and NK for HR, and NERC grant NE/N003942/1 to KRE.

Author contributions

KRE, NK and HR conceived the study. HR and KRE collected samples and designed the experiments. HR generated data, performed all analyses and drafted the manuscript. KRE, NK and HR all contributed to the writing of the final version of manuscript.

Conflict of interest

The authors declare no conflict of interest.

Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at <https://doi.org/10.1016/j.ympbev.2018.05.029>.

References

Alexander, D.H., Novembre, J., Lange, K., 2009. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.* 19, 1655–1664. <http://dx.doi.org/10.1101/gr.094052.109>.

Arrayago, M.J., Bea, A., Heulin, B., 1996. Hybridization experiment between oviparous and viviparous strains of *Lacerta vivipara*: A new insight into the evolution of viviparity in reptiles. *Herpetologica* 52, 333–342. <http://dx.doi.org/10.2307/3892653>.

Arribas, O.J., 2009. Morphological variability of the Cantabro-Pyrenean populations of *Zootoca vivipara* with description of a new subspecies. *Herpetozoa* 21, 123–146.

Ballard, J.W.O., Whitlock, M.C., 2004. The incomplete natural history of mitochondria. *Mol. Ecol.* 13, 729–744. <http://dx.doi.org/10.1046/j.1365-294X.2003.02063.x>.

Beaulieu, J.M., Oliver, J.C., O'Meara, O., 2012. corHMM: hidden Markov models in R, version 1.

Blackburn, D.G., 2015. Evolution of viviparity in squamate reptiles: Reversibility reconsidered. *J. Exp. Zool. Part B Mol. Dev. Evol.* 324, 473–486. <http://dx.doi.org/10.1002/jez.b.22625>.

Blackburn, D.G., 2006. Squamate reptiles as model organisms for the evolution of

viviparity. *Herpetol. Monogr.* 20, 131. [http://dx.doi.org/10.1655/0733-1347\(2007\)20\[131:SRAMOF\]2.0.CO;2](http://dx.doi.org/10.1655/0733-1347(2007)20[131:SRAMOF]2.0.CO;2).

Blackburn, D.G., 1999. Are viviparity and egg-guarding evolutionarily labile in squamates? *Herpetologica* 55, 556–573.

Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard, M.A., Rambaut, A., Drummond, A.J., 2014. BEAST 2: A Software platform for Bayesian evolutionary analysis. *PLoS Comput. Biol.* 10. <http://dx.doi.org/10.1371/journal.pcbi.1003537>.

Bryant, D., Bouckaert, R., Felsenstein, J., Rosenberg, N.A., Roychoudhury, A., 2012. Inferring species trees directly from biallelic genetic markers: Bypassing gene trees in a full coalescent analysis. *Mol. Biol. Evol.* 29, 1917–1932. <http://dx.doi.org/10.1093/molbev/mss086>.

Catchen, J.M., Amores, A., Hohenlohe, P., Cresko, W., Postlethwait, J.H., 2011. Stacks: Building and genotyping loci de novo from short-read sequences. *G3 Genes Genom. Genet.* 1, 171–182. <http://dx.doi.org/10.1534/g3.111.000240>.

Collin, R., Cipriani, R., 2003. Dollo's law and the re-evolution of shell coiling. *Proc. Biol. Sci.* 270, 2551–2555. <http://dx.doi.org/10.1098/rspb.2003.2517>.

Collin, R., Miglietta, M.P., 2008. Reversing opinions on Dollo's Law. *Trends Ecol. Evol.* 23, 602–609. <http://dx.doi.org/10.1016/j.tree.2008.06.013>.

Cornetti, L., Belluardo, F., Ghielmi, S., Giovine, G., Ficetola, G.F., Bertorelle, G., Vernesi, C., Haufler, H.C., 2015a. Reproductive isolation between oviparous and viviparous lineages of the Eurasian common lizard *Zootoca vivipara* in a contact zone. *Biol. J. Linn. Soc.* 114, 566–573. <http://dx.doi.org/10.1111/bj.12478>.

Cornetti, L., Ficetola, G.F., Hoban, S., Vernesi, C., 2015b. Genetic and ecological data reveal species boundaries between viviparous and oviparous lizard lineages. *Heredity* 115, 517–526. <http://dx.doi.org/10.1038/hdy.2015.54>.

Cornetti, L., Menegon, M., Giovine, G., Heulin, B., Vernesi, C., 2014. Mitochondrial and nuclear DNA survey of *Zootoca vivipara* across the eastern Italian Alps: Evolutionary relationships, historical demography and conservation implications. *PLoS One* 9. <http://dx.doi.org/10.1371/journal.pone.0085912>.

Darriba, D., Taboada, G.L., Doallo, R., Posada, D., 2012. jModelTest2: more models, new heuristics and parallel computing. *Nat. Methods* 9, 772. <http://dx.doi.org/10.1038/nmeth.2109>.

Delsuc, F., Brinkmann, H., Philippe, H., 2005. Phylogenomics and the reconstruction of the tree of life. *Nat. Rev. Genet.* 6, 361–375. <http://dx.doi.org/10.1038/nrg1603>.

Eaton, D.A.R., Spriggs, E.L., Park, B., Donoghue, M.J., 2017. Misconceptions on missing data in RAD-seq phylogenetics with a deep-scale example from flowering plants. *Syst. Biol.* 66, 399–412. <http://dx.doi.org/10.1093/sysbio/syw092>.

Fairbairn, J., Shine, R., Moritz, C., Frommer, M., 1998. Phylogenetic relationships between oviparous and viviparous populations of an Australian lizard (*Lerista bougainvillii*, Scincidae). *Mol. Phylogenet. Evol.* 10, 95–103. <http://dx.doi.org/10.1006/mpev.1997.0468>.

Freire, N.P., Tennant, M.R., Miyamoto, M.M., 2003. Microarray analysis of reptiles and amphibians: application in ecology and evolution. *Zool. Stud.* 42, 391–404.

Fuentes-Pardo, A.P., Ruzzante, D.E., 2017. Whole-genome sequencing approaches for conservation biology: Advantages, limitations and practical recommendations. *Mol. Ecol.* 26, 5369–5406. <http://dx.doi.org/10.1111/mec.14264>.

Giarla, T.C., Esselstyn, J.A., 2015. The challenges of resolving a rapid, recent radiation: empirical and simulated phylogenomics of philippine shrews. *Syst. Biol.* 64, 727–740. <http://dx.doi.org/10.1093/sysbio/syv029>.

Goldberg, E.E., Igić, B., 2008. On phylogenetic tests of irreversible evolution. *Evolution* 62, 2727–2741. <http://dx.doi.org/10.1111/j.1558-5646.2008.00505.x>.

Gould, S.J., 1970. Dollo's law: irreversibility and the status of evolutionary laws. *J. Hist. Biol.* 3, 189–212.

Griffith, O.W., Blackburn, D.G., Brandley, M.C., Van Dyke, J.U., Whittington, C.M., Thompson, M.B., 2015. Ancestral state reconstructions require biological evidence to test evolutionary hypotheses: A case study examining the evolution of reproductive mode in squamate reptiles. *J. Exp. Zool. Part B Mol. Dev. Evol.* 324, 493–503. <http://dx.doi.org/10.1002/jez.b.22614>.

Guillaume, C.P., Heulin, B., Pavlinov, I.Y., Semenov, D.V., Bea, A., Vogrin, N., Surget-Groba, Y., 2006. Morphological variations in the common lizard, *Lacerta (Zootoca) vivipara*. *Russ. J. Herpetol.* 13, 1–10.

Hahn, M.W., Nakhleh, L., 2016. Irrational exuberance for resolved species trees. *Evolution* 70, 7–17. <http://dx.doi.org/10.1111/evo.12832>.

Heulin, B., Ghielmi, S., Vogrin, N., Surget-Groba, Y., Guillaume, C.P., 2002. Variation in eggshell characteristics and in intrauterine egg retention between two oviparous clades of the lizard *Lacerta vivipara*: Insight into the oviparity-viviparity continuum in squamates. *J. Morphol.* 252, 255–262. <http://dx.doi.org/10.1002/jmor.1103>.

Huang, H., Lacey Knowles, L., 2016. Unforeseen consequences of excluding missing data from next-generation sequences: Simulation study of rad sequences. *Syst. Biol.* 65, 357–365. <http://dx.doi.org/10.1093/sysbio/syu046>.

Jiang, W., Chen, S.Y., Wang, H., Li, D.Z., Wiens, J.J., 2014. Should genes with missing data be excluded from phylogenetic analyses? *Mol. Phylogenet. Evol.* 80, 308–318. <http://dx.doi.org/10.1016/j.ympbev.2014.08.006>.

Kearse, M., Moir, R., Wilson, A., Stones-Havas, S., Cheung, M., Sturrock, S., Buxton, S., Cooper, A., Markowitz, S., Duran, C., Thierer, T., Ashton, B., Meintjes, P., Drummond, A., 2012. Geneious Basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28, 1647–1649. <http://dx.doi.org/10.1093/bioinformatics/bts199>.

Keenan, K., McGinnity, P., Cross, T.F., Crozier, W.W., Prodöhl, P.A., 2013. diveRsity: An R package for the estimation and exploration of population genetics parameters and their associated errors. *Methods Ecol. Evol.* 4, 782–788. <http://dx.doi.org/10.1111/2041-210X.12067>.

King, B., Lee, M.S.Y., 2015. Ancestral state reconstruction, rate heterogeneity, and the evolution of reptile viviparity. *Syst. Biol.* 64, 532–544. <http://dx.doi.org/10.1093/sysbio/syv005>.

- Kupriyanova, L.A., Mayer, W., Böhme, W., 2006. Karyotype diversity of the Eurasian lizard *Zootoca vivipara* (Jacquin, 1787) from Central Europe and the evolution of viviparity, *Herpetologia Bonnensis* II. In: Proceedings of the 13th Congress of the Societas Europaea Herpetologica. Bonn.
- Kupriyanova, L., Kuksin, A., Odierna, G., 2008. Karyotype, chromosome structure, reproductive modalities of three Southern Eurasian populations of the common lacertid lizard, *Zootoca vivipara* (Jacquin, 1787). *Acta Herpetol.* 3, 99–106.
- Le Galliard, J.F., Le Bris, M., Clobert, J., 2003. Timing of locomotor impairment and shift in thermal preferences during gravidity in a viviparous lizard. *Funct. Ecol.* 17, 877–885. <http://dx.doi.org/10.1046/j.0269-8463.2003.00800.x>.
- Leavitt, D.H., Marion, A.B., Hollingsworth, B.D., Reeder, T.W., 2017. Multilocus phylogeny of alligator lizards (*Elgaria*, Anguillidae): Testing mtDNA introgression as the source of discordant molecular phylogenetic hypotheses. *Mol. Phylogenet. Evol.* 110, 104–121. <http://dx.doi.org/10.1016/j.ympev.2017.02.010>.
- Leavitt, S.D., Grewe, F., Widhelm, T., Muggia, L., Wray, B., Lumsch, H.T., 2016. Resolving evolutionary relationships in lichen-forming fungi using diverse phylogenomic datasets and analytical approaches. *Sci. Rep.* 6, 22262. <http://dx.doi.org/10.1038/srep22262>.
- Lee, M.S.Y., Shine, R., 1998. Reptilian viviparity and Dollo's law. *Evolution* 52, 1441–1450.
- Li, H., Durbin, R., 2010. Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics* 26, 589–595. <http://dx.doi.org/10.1093/bioinformatics/btp698>.
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., Durbin, R., 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25, 2078–2079. <http://dx.doi.org/10.1093/bioinformatics/btp352>.
- Lindtke, D., Mayer, W., Böhme, W., 2010. Identification of a contact zone between oviparous and viviparous common lizards (*Zootoca vivipara*) in central Europe: Reproductive strategies and natural hybridization. *Salamandra* 46, 73–82.
- Lynch, V.J., Wagner, G.P., 2010. Did egg-laying boas break Dollo's law? Phylogenetic evidence for reversal to oviparity in sand boas (*Eryx*: Boidae). *Evolution* 64, 207–216. <http://dx.doi.org/10.1111/j.1558-5646.2009.00790.x>.
- Mastretta-Yanes, A., Arrigo, N., Alvarez, N., Jorgensen, T.H., Piñero, D., Emerson, B.C., 2015. Restriction site-associated DNA sequencing, genotyping error estimation and *de novo* assembly optimization for population genetic inference. *Mol. Ecol. Resour.* 15, 28–41. <http://dx.doi.org/10.1111/1755-0998.12291>.
- Mayer, W., Böhme, W., Tiedemann, F., Bischoff, W., 2000. On oviparous populations of *Zootoca vivipara* (Jacquin, 1787) in south-eastern Central Europe and their phylogenetic relationship to neighbouring viviparous and South-west European oviparous populations. *Herpetozoa* 13, 59–69.
- Miller, M.A., Pfeiffer, W., Schwartz, T., 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: 2010 Gateway Computing Environments Workshop, GCE 2010. doi:10.1109/GCE.2010.5676129.
- Mirarab, S., Reaz, R., Bayzid, M.S., Zimmermann, T.S., Swenson, M., Warnow, T., 2014. ASTRAL: Genome-scale coalescent-based species tree estimation. *Bioinformatics*. <http://dx.doi.org/10.1093/bioinformatics/btu462>.
- Murphy, B.F., Thompson, M.B., 2011. A review of the evolution of viviparity in squamate reptiles: The past, present and future role of molecular biology and genomics. *J. Comp. Physiol. B Biochem. Syst. Environ. Physiol.* 181, 575–594. <http://dx.doi.org/10.1007/s00360-011-0584-0>.
- Near, T.J., Keck, B.P., 2013. Free from mitochondrial DNA: Nuclear genes and the inference of species trees among closely related darter lineages (Teleostei: Percidae: Etheostomatinae). *Mol. Phylogenet. Evol.* 66, 868–876. <http://dx.doi.org/10.1016/j.ympev.2012.11.009>.
- Odierna, G., Aprea, G., Capriglione, T., Arribas, O.J., Kupriyanova, L.A., Olmo, E., 1998. Progressive differentiation of the W sex-chromosome between oviparous and viviparous populations of *Zootoca vivipara* (Reptilia, Lacertidae). *Ital. J. Zool.* 65, 295–302. <http://dx.doi.org/10.1080/1125008809386761>.
- Odierna, G., Aprea, G., Capriglione, T., Puky, M., 2004. Chromosomal evidence for the double origin of viviparity in the European common lizard, *Lacerta* (*Zootoca*) *vivipara*. *Herpetol. J.* 14, 157–160.
- Pavlicev, M., Mayer, W., 2009. Fast radiation of the subfamily Lacertinae (Reptilia: Lacertidae): History or methodical artefact? *Mol. Phylogenet. Evol.* 52, 727–734. <http://dx.doi.org/10.1016/j.ympev.2009.04.020>.
- Peterson, B.K., Weber, J.N., Kay, E.H., Fisher, H.S., Hoekstra, H.E., et al., 2012. Double digest RADseq: an inexpensive method for *de novo* SNP discovery and genotyping in model and non-model species. *PLoS One* 7, 1–11. <http://dx.doi.org/10.1371/journal.pone.0037135>.
- Pickrell, J.K., Pritchard, J.K., 2012. Inference of population splits and mixtures from genome-wide allele frequency data. *PLoS Genet.* 8. <http://dx.doi.org/10.1371/journal.pgen.1002967>.
- Posada, D., 2016. Phylogenomics for systematic biology. *Syst. Biol.* 65, 353–356. <http://dx.doi.org/10.1093/sysbio/syw027>.
- Pyron, R.A., Burbrink, F.T., 2014. Early origin of viviparity and multiple reversions to oviparity in squamate reptiles. *Ecol. Lett.* 17, 13–21. <http://dx.doi.org/10.1111/ele.12168>.
- Pyron, R.A., Burbrink, F.T., Wiens, J.J., 2013. A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evol. Biol.* 13, 93. <http://dx.doi.org/10.1186/1471-2148-13-93>.
- Racimo, F., Sankararaman, S., Nielsen, R., Huerta-Sánchez, E., 2015. Evidence for archaic adaptive introgression in humans. *Nat. Rev. Genet.* 16, 359–371. <http://dx.doi.org/10.1038/nrg3936>.
- Rambaut, A., Drummond, A.J., 2009. Tracer V1.5. Available from <http://beast.bio.ed.ac.uk/Tracer>.
- Recknagel, H., Elmer, K.R., Meyer, A., 2013. A hybrid genetic linkage map of two ecologically and morphologically divergent Midas Cichlid fishes (*Amphilophus* spp.) obtained by massively parallel DNA sequencing (ddRADSeq). *G3 Genes Genom. Genet.* 3, 65–74. <http://dx.doi.org/10.1534/g3.112.003897>.
- Recknagel, H., Jacobs, A., Herzyk, P., Elmer, K.R., 2015. Double-digest RAD sequencing using Ion Proton semiconductor platform (ddRADseq-ion) with nonmodel organisms. *Mol. Ecol. Resour.* 15, 1316–1329. <http://dx.doi.org/10.1111/1755-0998.12406>.
- Rodríguez-Prieto, A., Giovine, G., Laddaga, L., Ghielmi, S., Cornetti, L., 2017. Very similar, but not identical: morphological taxonomic identification to improve the resolution of fine-scale distribution of *Zootoca* (*vivipara*) *caroliola*. *Amphibia-Reptilia*. <http://dx.doi.org/10.1163/15685381-00003120>.
- Rodríguez, A., Burgon, J.D., Lyra, M., Irisarri, I., Baurain, D., Blaustein, L., Göçmen, B., Künzel, S., Mable, B.K., Nolte, A.W., Veith, M., Steinfartz, S., Elmer, K.R., Philippe, H., Vences, M., 2017. Inferring the shallow phylogeny of true salamanders (*Salamandra*) by multiple phylogenomic approaches. *Mol. Phylogenet. Evol.* 115, 16–26. <http://dx.doi.org/10.1016/j.ympev.2017.07.009>.
- Rowe, H.C., Renaut, S., Guggisberg, A., 2011. RAD in the realm of next-generation sequencing technologies. *Mol. Ecol.* 20, 3499–3502. <http://dx.doi.org/10.1111/j.1365-294X.2011.05197.x>.
- Schmidler, J.F., Böhme, W., 2011. Synonymy and nomenclatural history of the common or viviparous lizard, by this time: *Zootoca vivipara* (Lichtenstein, 1823). *Bonn Zool. Bull.* 60, 214–228.
- Shafer, A.B.A., Peart, C.R., Tusso, S., Maayan, I., Brelford, A., Wheat, C.W., Wolf, J.B.W., 2017. Bioinformatic processing of RAD-seq data dramatically impacts downstream population genetic inference. *Methods Ecol. Evol.* 8, 907–917. <http://dx.doi.org/10.1111/2041-210X.12700>.
- Shimodaira, H., 2002. An approximately unbiased test of phylogenetic tree selection. *Syst. Biol.* 51, 492–508. <http://dx.doi.org/10.1080/10635150290069913>.
- Shimodaira, H., Hasegawa, M., 2001. CONSEL: for assessing the confidence of phylogenetic tree selection. *Bioinformatics* 17, 1246–1247. <http://dx.doi.org/10.1093/bioinformatics/17.12.1246>.
- Shimodaira, H., Hasegawa, M., 1999. Multiple comparisons of log-likelihoods with applications to phylogenetic inference. *Mol. Biol. Evol.* 16, 1114–1116. <http://dx.doi.org/10.1093/oxfordjournals.molbev.a026201>.
- Shine, R., Lee, M.S.Y., 1999. A reanalysis of the evolution of viviparity and egg-guarding in squamate reptiles. *Herpetologica* 55, 538–549.
- Sites, J.W., Reeder, T.W., Wiens, J.J., 2011. Phylogenetic insights on evolutionary novelties in lizards and snakes: sex, birth, bodies, niches, and venom. *Annu. Rev. Ecol. Syst.* 42, 227–244. <http://dx.doi.org/10.1146/annurev-ecolsys-102710-145051>.
- Smith, M.F., Patton, J.L., 1991. Variation in mitochondrial cytochrome b sequence in natural populations of South American akodontine rodents (Muridae: Sigmodontinae). *Mol. Biol. Evol.* 8, 85–103. <http://dx.doi.org/10.1093/oxfordjournals.molbev.a040638>.
- Stamatakis, A., 2006. RAxML 7.0.4 Manual. *Bioinformatics* 22 (21), 2688–2690. <http://dx.doi.org/10.1093/bioinformatics/btl446>.
- Stern, D.L., 2013. The genetic causes of convergent evolution. *Nat. Rev. Genet.* 14, 751–764. <http://dx.doi.org/10.1038/nrg3483>.
- Streicher, J.W., Schulte, J.A., Wiens, J.J., 2016. How should genes and taxa be sampled for phylogenomic analyses with missing data? An empirical study in Iguanian lizards. *Syst. Biol.* 65, 128–145. <http://dx.doi.org/10.1093/sysbio/syw058>.
- Surget-Groba, Y., Heulin, B., Guillaume, C.-P., Thorpe, R.S., Kupriyanova, L., Vogrin, N., Maslak, R., Mazzotti, S., Venczel, M., Ghira, I., Odierna, G., Leontyeva, O., Monney, J.C., Smith, N., 2001. Intraspecific phylogeography of *Lacerta vivipara* and the evolution of viviparity. *Mol. Phylogenet. Evol.* 18, 449–459. <http://dx.doi.org/10.1006/mpev.2000.0896>.
- Surget-Groba, Y., Heulin, B., Guillaume, C.P., Puky, M., Semenov, D., Orlova, V., Kupriyanova, L., Ghira, I., Smajda, B., 2006. Multiple origins of viviparity, or reversal from viviparity to oviparity? The European common lizard (*Zootoca vivipara*, Lacertidae) and the evolution of parity. *Biol. J. Linn. Soc.* 87, 1–11. <http://dx.doi.org/10.1111/j.1095-8312.2006.00552.x>.
- Swofford, D.L., 2002. Phylogenetic analysis using parsimony. *Options* 42, 294–307. <http://dx.doi.org/10.1007/BF02198856>.
- Velekei, B., Lakatos, F., Covaciu-Marcov, S.D., Sas-Kovács, I., Puky, M., 2015. New *Zootoca vivipara* (Lichtenstein, 1823) haplogroup in the Carpathians. *North. West. J. Zool.*
- Wallis, G.P., Cameron-Christie, S.R., Kennedy, H.L., Palmer, G., Sanders, T.R., Winter, D.J., 2017. Interspecific hybridization causes long-term phylogenetic discordance between nuclear and mitochondrial genomes in freshwater fishes. *Mol. Ecol.* 26, 3116–3127. <http://dx.doi.org/10.1111/mec.14096>.
- Whiting, M.F., Bradler, S., Maxwell, T., 2003. Loss and recovery of wings in stick insects. *Nature* 421, 264–267. <http://dx.doi.org/10.1038/nature01274.1>.
- Wiens, J.J., 2011. Re-evolution of lost mandibular teeth in frogs after more than 200 million years, and re-evaluating dollo's law. *Evolution* 65, 1283–1296. <http://dx.doi.org/10.1111/j.1558-5646.2011.01221.x>.
- Willis, S.C., Farias, I.P., Ortí, G., 2014. Testing mitochondrial capture and deep coalescence in Amazonian cichlid fishes (Cichlidae: *Cichla*). *Evolution* 68, 256–268. <http://dx.doi.org/10.1111/evo.12230>.
- Wright, A.M., Lyons, K.M., Brandley, M.C., Hillis, D.M., 2015. Which came first: The lizard or the egg? Robustness in phylogenetic reconstruction of ancestral states. *J. Exp. Zool. Part B Mol. Dev. Evol.* 324, 504–516. <http://dx.doi.org/10.1002/jez.b.22642>.